

**Total Synthesis of Dendroamide A:  
Oxazole and Thiazole Construction Using  
an Oxodiphosphonium Salt**

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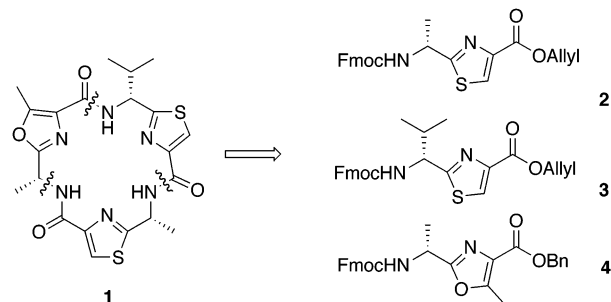
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**Abstract:** The total synthesis of dendroamide A (**1**), a multidrug-resistance reversing bistratamide-type peptide-derived macrocycle, has been accomplished in 19% yield. Fmoc-protected amino acids were condensed into appropriately protected dipeptides which were treated with bis-(triphenyl)oxodiphosphonium trifluoromethanesulfonate to afford oxazoles and thiazolines (oxidized to thiazoles) with high chemo- and stereoselectivity. The convergent condensation of three heterocyclic amino acids followed by macrocyclization afforded the natural product.

Dendroamide A (**1**) was first isolated from the terrestrial blue-green alga (cyanobacterium) *Stigonema dendroideum fremy* in 1996.<sup>1</sup> This natural product reverses multiple drug resistance (MDR)<sup>2</sup> by acting as a P-glycoprotein and MRP1 antagonist at noncytotoxic doses. Dendroamide A could be useful against cancers exhibiting MDR and therefore has attracted the attention of synthetic and natural product chemists. Its total synthesis has been reported by two groups.<sup>3</sup> The thiazole substructure was prepared either by Hantzsch's procedure using a thioamide as an intermediate<sup>4</sup> or by employing a condensation reaction between cysteine esters and *N*-protected imino esters, followed by the oxidation of the resulting thiazolines.<sup>5</sup> Synthesis of the oxazole substructure was achieved by oxidation of the oxazoline, made from a  $\beta$ -hydroxy amide utilizing the Burgess reagent.<sup>6</sup>

In this paper, we report the synthesis of dendroamide A in 19% overall yield. Highlights include the preparation



**FIGURE 1.** Retrosynthetic analysis of dendroamide A (**1**).

of the oxazole substructure from a  $\beta$ -ketodipeptide and the thiazolines from fully protected cysteine-containing dipeptides, in all cases utilizing bis(triphenyl)oxodiphosphonium trifluoromethanesulfonate.<sup>7</sup>

Our retrosynthetic analysis of dendroamide A (**1**), outlined in Figure 1, features disconnection at three amide bonds. The resulting appropriately protected heterocyclic amino acid fragments **2–4** are derived from dipeptides, which were synthesized from ordinary Fmoc-protected amino acids. The ketone side chain used to produce **4** results from the Dess–Martin oxidation of a threonine-containing dipeptide.

As shown in Scheme 1, compound **5** was synthesized by coupling *N*-Fmoc-L-S-tritylcysteine with allyl alcohol utilizing HBTU (2-(1*H*-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate) and HOBt (*N*-hydroxybenzotriazole) in the presence of DIEA (*N,N*-diisopropylethylamine). Diethylamine removal of the Fmoc protecting group afforded an amine that was coupled with *N*-Fmoc-D-alanine or *N*-Fmoc-D-valine employing HBTU and HOBt in the presence of DIEA, yielding dipeptides **6** and **7**, respectively. Transformation of the fully protected cysteine-containing dipeptides to thiazolines **8** and **9** was accomplished in high yield utilizing bis(triphenyl)oxodiphosphonium trifluoromethanesulfonate,<sup>7,8</sup> generated from triphenyl phosphine oxide and triflic anhydride. The thiazolines in **8** and **9** were oxidized to thiazoles using activated manganese oxide. The stereochemical purity of **2** and **3** exceeded 96% ee as ascertained by chiral HPLC.

The synthesis of oxazole fragment **4** is outlined in Scheme 2. Dipeptide **10** was synthesized by coupling Fmoc-D-alanine and L-threonine benzyl ester. A Dess–Martin oxidization of the hydroxy group in dipeptide **10** led to ketone **11** in 80% yield.<sup>9</sup> Bis(triphenyl)oxodiphosphonium trifluoromethanesulfonate was utilized to convert the  $\beta$ -ketodipeptide **11** to the protected oxazole amino acid **4** in 84% yield without compromising the Aladerived stereocenter (98% ee). Unlike the oxobisphosphonium salt utilized here, the PPh<sub>3</sub>I<sub>2</sub> reagent utilized

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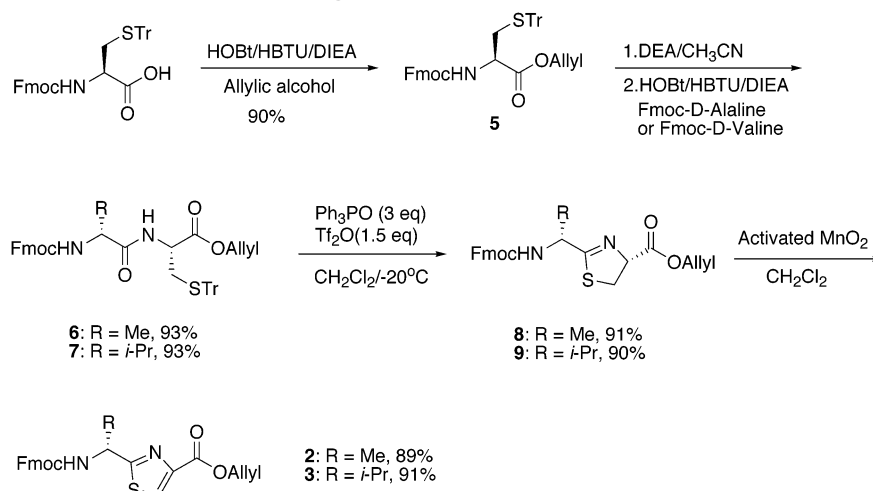
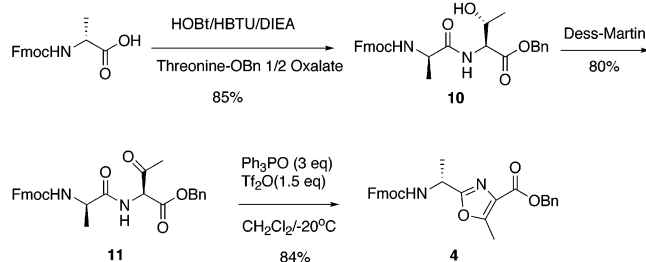
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SCHEME 1. Synthesis of Thiazole-Containing Amino Acids **2** and **3**SCHEME 2. Synthesis of Oxazole-Containing Amino Acid **4** from a Thr-Based Dipeptide

previously for this transformation requires the use of a base increasing the risk of racemization.<sup>10</sup>

The allyl group in compound **3** was removed utilizing a palladium catalyst, generated from Pd(OAc)<sub>2</sub> and polymer-supported triphenylphosphine, in the presence of phenylsilane.<sup>11</sup> The use of this solid-phase catalyst greatly simplified workup of the reaction, as the carboxylic acid could be separated from the catalyst by filtration through a short silica gel column. The Fmoc group in compound **2** was removed by diethylamine, allowing its amino terminus to be coupled with the carboxylic acid derived from **3**, utilizing HBTU and HOBT in the presence of DIEA, affording the peptidic bis-heterocycle **12** (92%). The benzyl group in **4** was removed by a Pd/C-mediated hydrogenation. The resulting carboxylic acid was coupled with the free amine resulting from the diethylamine deprotection of compound **12**, yielding compound **13** in 94% yield. The macrolide precursor **14** can be obtained from peptide **13** by removing the Fmoc and allyl groups as described above. The final cyclization promoted by PyBOP (benzotriazole-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate) and DMAP can be achieved by slowly adding **14** to a solution of PyBOP and DMAP in CH<sub>2</sub>Cl<sub>2</sub>/DMF (2/1) with a syringe pump affording **1** in 81% yield as a white solid (Scheme 3). The <sup>1</sup>H and <sup>13</sup>C NMR spectra of **1** are identical to those reported in the literature.<sup>1, 3</sup>

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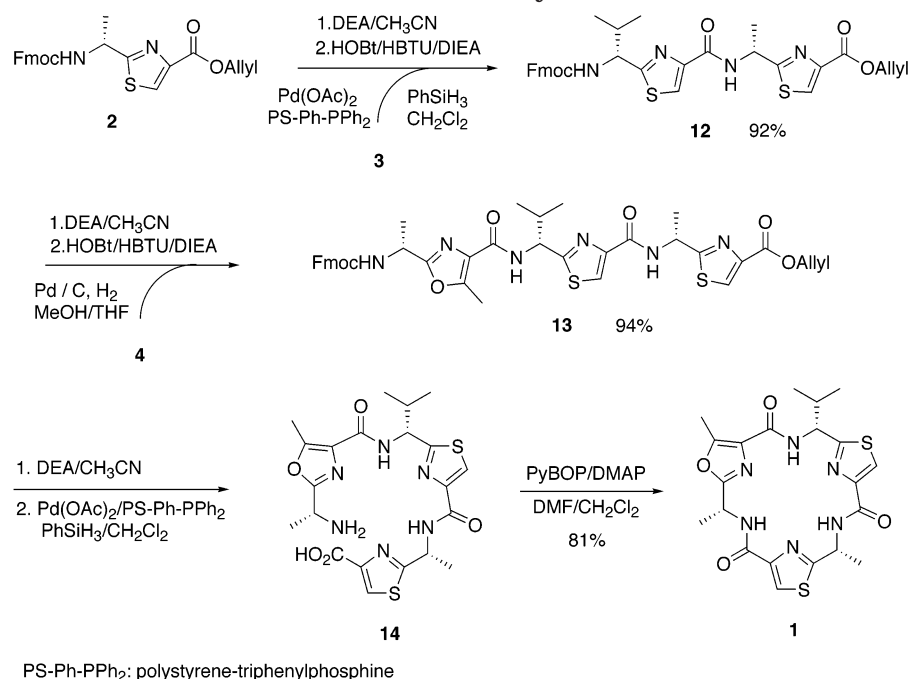
In summary, dendroamide **A** (**1**) has been synthesized in a highly convergent fashion from three appropriately protected heterocyclic amino acids. The oxazole amino acid results from cyclodehydration of a β-ketodipeptide. The thiazole amino acids are obtained from oxidation of their corresponding thiazoline amino acids, which are synthesized by the cyclodehydration of fully protected cysteine-based dipeptides. In each case bis(triphenyl)-oxodiphosphonium trifluoromethanesulfonate was employed to make the heterocyclic amino acids from the dipeptide by amide carbonyl activation/dehydration. The oxophosphonium salt accomplished these reactions with notable chemo- and stereoselectivity. The final macrocyclization proceeds in high yield (81%) affording **1** in an overall yield of 19%.

## Experimental Section

**Compound 5.** A solution of *N*<sup>ε</sup>-Fmoc-L-Cys(S-trityl)-OH (5.86 g, 10 mmol) in DMF (50 mL) was treated with HOBT·H<sub>2</sub>O (1.68 g, 11 mmol) and HBTU (4.17 g, 11 mmol) at 25 °C. After 10 min, allyl alcohol (1.36 mL, 20 mmol) and DIEA (3.65 mL, 21 mmol) were added separately, and the resulting mixture was stirred at 25 °C overnight. After regular workup, the resulting crude product was purified by flash chromatography (EtOAc/hexanes = 1/3) to afford compound **5** (5.63 g, 90%) as a white foam: [α]<sub>D</sub><sup>25</sup> = +18.1 (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 2.63–2.70 (m, 2H), 4.22 (dd, *J* = 7.0, 6.6 Hz, 1H), 4.33–4.39 (m, 3H), 4.58 (m, 2H), 5.23 (d, *J* = 10.5 Hz, 1H), 5.28–5.30 (m, 2H), 5.86 (m, 1H), 7.18–7.29 (m, 11H), 7.37–7.40 (m, 8H), 7.60 (dd, *J* = 6.1, 5.7 Hz, 2H), 7.75 (dd, *J* = 6.6, 6.1 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 34.0, 47.0, 52.9, 66.2, 67.0, 67.1, 118.7, 119.9, 125.1 (2 C), 126.8, 126.9, 127.0, 127.7, 127.9, 128.0 (3 C), 129.4 (3 C), 129.5, 131.3, 141.2 (2 C), 143.7, 143.8, 144.2, 155.5, 170.2; HRMS (MALDI-FTMS) calcd for C<sub>40</sub>H<sub>35</sub>NO<sub>4</sub>S (M + Na<sup>+</sup>) 648.2179, found 648.2176.

**Compound 6 (General Procedure for Diethylamine-Mediated Deprotection of Fmoc Group and HOBT, HBTU-Mediated Peptide Coupling).** Diethylamine (30 mL) was added to a solution of **5** (5.63 g, 9 mmol) in CH<sub>3</sub>CN (30 mL), and the resulting mixture was stirred at 25 °C for 30 min to ensure complete removal of the Fmoc protecting group. After concentration in vacuo, the reaction mixture was azeotroped to dryness with CH<sub>3</sub>CN (2 × 30 mL), and the residue was dissolved in DMF (40 mL). In another flask, a solution of *N*<sup>ε</sup>-Fmoc-D-alanine (3.11 g, 10 mmol) in DMF (40 mL) was treated with HOBT·H<sub>2</sub>O (1.53 g, 10 mmol) and HBTU (3.79 g, 10 mmol). After 10 min, this mixture and DIEA (3.65 mL, 21 mmol) were sequentially added to the above free amino ester. The reaction

## SCHEME 3. Synthesis of Dendroamide A (1) from 3 Heterocyclic Amino Acids Derived from Dipeptides



was stirred at 25 °C for 8 h. After regular workup, the resulting crude product was purified by flash chromatography (EtOAc/hexanes = 1/3) to afford compound **6** (5.83 g, 93%) as a pale oil:  $[\alpha]_D^{24} = +42.7$  (*c* 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 1.40 (d, *J* = 6.6 Hz, 3H), 2.61–2.63 (m, 2H), 4.18 (t, *J* = 7.0 Hz, 1H), 4.30–4.38 (m, 3H), 4.52 (m, 1H), 4.56 (d, *J* = 5.7 Hz, 2H), 5.20 (d, *J* = 10.5 Hz, 1H), 5.25 (dd, *J* = 17.1, 1.3 Hz, 1H), 5.48 (d, *J* = 7.0 Hz, 1H), 5.85–5.79 (m, 1H), 6.67 (d, *J* = 6.6 Hz, 1H), 7.18–7.29 (m, 11H), 7.35–7.39 (m, 8H), 7.54 (d, *J* = 7.5 Hz, 1H), 7.56 (d, *J* = 7.0 Hz, 1H), 7.74 (d, *J* = 7.0 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 18.8, 33.7, 47.0, 50.3, 51.2, 66.2, 66.7, 67.1, 118.8, 119.9, 125.0, 125.1, 126.8, 127.0 (2 C), 127.6, 127.8, 128.0, 129.4, 131.3, 141.2, 143.6, 143.9, 144.1, 155.8, 169.8, 171.9; HRMS (MALDI-FTMS) calcd for C<sub>43</sub>H<sub>40</sub>N<sub>2</sub>O<sub>5</sub>S (M + Na<sup>+</sup>) 719.2550, found 719.2551.

**Compound 7.** Compound **7** was synthesized from **5** and *N*<sup>ε</sup>-Fmoc-D-valine in 93% yield as a white foam by following the procedure used for the synthesis of **6**:  $[\alpha]_D^{24} = +33.5$  (*c* 0.68, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 0.93 (d, *J* = 6.6 Hz, 3H), 0.97 (d, *J* = 5.7 Hz, 3H), 2.14 (m, 1H), 2.61 (m, 2H), 4.07 (m, 1H), 4.19 (m, 1H), 4.36 (m, 2H), 4.56 (m, 1H), 4.60 (m, 2H), 5.23 (d, *J* = 10.5 Hz, 1H), 5.28 (d, *J* = 17.1 Hz, 1H), 5.37 (d, *J* = 8.8 Hz, 1H), 5.85 (m, 1H), 6.42 (d, *J* = 7.9 Hz, 1H), 7.19–7.30 (m, 11H), 7.36–7.39 (m, 8H), 7.57 (m, 2H), 7.76 (d, *J* = 6.6 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 17.6, 19.2, 31.2, 33.8, 47.1, 51.2, 59.9, 66.3, 66.7, 67.0, 118.8, 119.9, 125.1, 126.8, 126.9, 127.0, 127.6, 127.9, 128.0, 129.3, 129.4, 131.3, 141.2, 143.6, 143.9, 144.1, 156.3, 169.8, 170.8; HRMS (MALDI-FTMS) calcd for C<sub>45</sub>H<sub>44</sub>N<sub>2</sub>O<sub>5</sub>S (M + Na<sup>+</sup>) 747.2863, found 747.2854.

**Compound 8.** To a solution of triphenylphosphine oxide (8.35 g, 30 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (100 mL) was added trifluoromethanesulfonic anhydride (2.35 mL, 15 mmol) slowly at 0 °C. The reaction mixture was stirred for 10 min at 0 °C and then adjusted to –20 °C using a brine–ice bath. Then, a solution of **6** (6.97 g, 10 mmol) in 10 mL CH<sub>2</sub>Cl<sub>2</sub> was added. The reaction progress was monitored by TLC and was completed in 2 h. After regular workup, the resulting crude product was purified by flash chromatography (EtOAc/hexanes = 1/3) to afford compound **8** (3.97 g, 91%) as an oil:  $[\alpha]_D^{24} = +37.8$  (*c* 2.67, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 1.46 (d, *J* = 7.0 Hz, 3H), 3.51 (t, *J* = 11.0 Hz, 1H), 3.59 (dd, *J* = 11.0, 8.0 Hz, 1H), 4.21 (t, *J* = 7.0 Hz, 1H), 4.34 (dd, *J* = 10.1, 7.5 Hz, 1H), 4.42 (dt, *J* = 10.5, 7.0 Hz, 1H), 4.67–4.69 (m, 3H), 5.12 (dd, *J* = 9.2, 8.8 Hz, 1H), 5.23 (d, *J* = 10.5 Hz, 1H), 5.33 (d, *J* = 17.1 Hz, 1H), 5.66

(d, *J* = 7.5 Hz, 1H), 5.91 (m, 1H), 7.26–7.30 (m, 2H), 7.38 (t, *J* = 7.5 Hz, 2H), 7.58 (d, *J* = 7.5 Hz, 1H), 7.60 (d, *J* = 7.5 Hz, 1H), 7.74 (d, *J* = 7.5 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 20.2, 35.3, 47.0, 49.7, 66.1, 66.7, 77.7, 118.8, 119.8, 124.9, 125.0, 126.9, 127.0, 127.6, 127.7, 127.8, 131.3, 141.1 (2 C), 143.6, 143.8, 146.8, 155.4, 170.0, 176.9; HRMS (MALDI-FTMS) calcd for C<sub>24</sub>H<sub>24</sub>N<sub>2</sub>O<sub>4</sub>S (M + H<sup>+</sup>) 437.1529, found 437.1525.

**Compound 9.** Compound **9** was synthesized from **7** in 90% yield as a white solid by following the procedure used for the synthesis of **8**: mp 73–75 °C;  $[\alpha]_D^{24} = +47.3$  (*c* 1.02, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 0.94 (d, *J* = 7.0 Hz, 3H), 1.01 (d, *J* = 6.6 Hz, 3H), 2.19 (m, 1H), 3.54 (dd, *J* = 10.0, 9.6 Hz, 1H), 3.62 (m, 1H), 4.25 (dd, *J* = 7.5, 7.0 Hz, 1H), 4.38 (m, 1H), 4.44 (m, 1H), 4.56 (dd, *J* = 8.8, 5.3 Hz, 1H), 4.68 (d, *J* = 6.1 Hz, 2H), 5.17 (dd, *J* = 9.2, 8.3 Hz, 1H), 5.24 (d, *J* = 9.2 Hz, 1H), 5.32–5.35 (m, 1H), 5.42 (d, *J* = 8.8 Hz, 1H), 5.91 (m, 1H), 7.27–7.33 (m, 2H), 7.40 (dd, *J* = 7.5, 7.0 Hz, 2H), 7.60 (d, *J* = 7.5 Hz, 1H), 7.61 (d, *J* = 7.5 Hz, 1H), 7.76 (d, *J* = 7.9 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 17.0, 19.3, 32.2, 35.3, 47.1, 58.8, 66.2, 66.9, 77.8, 118.9, 119.9, 125.0, 125.1, 127.0, 127.1, 127.6, 127.8, 127.9, 131.4, 141.2, 143.7, 143.9, 146.8, 156.1, 170.1, 175.9; HRMS (MALDI-FTMS) calcd for C<sub>26</sub>H<sub>28</sub>N<sub>2</sub>O<sub>4</sub>S (M + H<sup>+</sup>) 465.1842, found 465.1846.

**Compound 2.** To a solution of **8** (436 mg, 1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL), activated MnO<sub>2</sub> (<5 μm, 85%, 1.02 g, 10 mmol) was added. The reaction mixture was stirred overnight at 25 °C, then filtered through a short silica gel and Celite column and washed with EtOAc. The organic solution was concentrated. The resulting crude product was purified by flash chromatography (EtOAc/hexanes = 1/3) to afford compound **2** (387 mg, 89%) as a white solid: mp 154–157 °C;  $[\alpha]_D^{24} = +19.8$  (*c* 0.61, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 1.63 (d, *J* = 5.7 Hz, 3H), 4.20 (m, 1H), 4.40 (m, 1H), 4.47 (dd, *J* = 10.5, 6.6 Hz, 1H), 4.84 (d, *J* = 4.8 Hz, 2H), 5.17 (m, 1H), 5.29 (d, *J* = 10.5 Hz, 1H), 5.40 (d, *J* = 17.1 Hz, 1H), 5.67 (br, 1H), 6.02 (m, 1H), 7.25–7.27 (m, 4H), 7.56 (br, 2H), 7.74 (br, 2H), 8.11 (s, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 21.6, 47.0, 49.2, 65.9, 66.8, 118.9, 119.9, 124.9, 125.0, 127.0, 127.5, 127.6, 131.7, 141.2, 143.6, 146.7, 155.5, 160.8, 173.9; HRMS (MALDI-FTMS) calcd for C<sub>24</sub>H<sub>22</sub>N<sub>2</sub>O<sub>4</sub>S (M + H<sup>+</sup>) 435.1373, found 435.1371; HPLC (Chiralcel OD-H column, 254 nm, 90/10 hexane/2-propanol to 70/30 hexane/2-propanol gradient over 60 min, flow = 1.0 mL/min) *t*<sub>R</sub> = 50.8 (S), 57.6 (R) min.

**Compound 3.** Compound **3** was synthesized from **9** in 91% yield as a white solid by following the procedure used for the

synthesis of **2**: mp 104–106 °C;  $[\alpha]^{24}_D = +42.7$  (*c* 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 0.94 (d, *J* = 7.0 Hz, 3H), 0.96 (d, *J* = 7.0 Hz, 3H), 2.43 (m, 1H), 4.22 (dd, *J* = 6.6, 6.1 Hz, 1H), 4.44 (m, 2H), 4.85 (d, *J* = 5.7 Hz, 2H), 4.94 (dd, *J* = 8.8, 6.6 Hz, 1H), 5.30 (d, *J* = 10.5 Hz, 1H), 5.41 (d, *J* = 15.1 Hz, 1H), 5.59 (d, *J* = 8.3 Hz, 1H), 6.04 (m, 1H), 7.31 (m, 2H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.60 (dd, *J* = 6.1, 5.3 Hz, 2H), 7.76 (d, *J* = 7.5 Hz, 2H), 8.10 (s, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 17.5, 19.4, 33.3, 47.1, 58.5, 65.9, 66.8, 118.8, 119.9, 124.9, 125.0, 127.0, 127.2, 127.6, 131.8, 141.2, 143.6, 143.7, 146.9, 156.0, 160.8, 172.2; HRMS (MALDI-FTMS) calcd for C<sub>26</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub>S (M + H<sup>+</sup>) 463.1686, found 463.1684; HPLC (Chiralcel OD-H column, 254 nm, 90/10 hexane/2-propanol to 70/30 hexane/2-propanol gradient over 60 min, flow = 1.0 mL/min) *t*<sub>R</sub> = 26.5 (*S*), 30.9 (*R*) min.

**Compound 10.** Compound **10** was synthesized from *N*<sup>ε</sup>-Fmoc-D-valine and L-threonine benzyl ester oxalate in 85% yield as a white foam by following the procedure used for the synthesis of **6**:  $[\alpha]^{24}_D = -25.7$  (*c* 0.77, DMSO); <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>, 25 °C) δ 1.04 (d, *J* = 6.1 Hz, 3H), 1.26 (d, *J* = 7.0 Hz, 3H), 4.18–4.27 (m, 5H), 4.37 (d, *J* = 8.8 Hz, 1H), 5.08–5.11 (m, 1H), 5.13–5.16 (m, 2H), 7.30–7.37 (m, 7H), 7.41 (t, *J* = 7.5 Hz, 2H), 7.62 (d, *J* = 7.5 Hz, 1H), 7.73 (m, 2H), 7.89–7.91 (m, 3H); <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 18.5, 20.1, 46.6, 57.6, 65.7, 65.9, 66.4, 120.1, 125.3 (2 C), 127.1, 127.6, 127.9, 128.3, 135.9, 140.7, 143.8, 143.9, 155.7, 170.4, 173.2; HRMS (MALDI-FTMS) calcd for C<sub>29</sub>H<sub>30</sub>N<sub>2</sub>O<sub>6</sub> (M + Na<sup>+</sup>) 525.1996, found 525.1996.

**Compound 11.** Dess–Martin periodinane (306 mg, 97% purity, 0.7 mmol) was added to a flask containing **10** (251 mg, 0.5 mmol) in 30 mL of CH<sub>2</sub>Cl<sub>2</sub>. The resulting reaction mixture was stirred at 25 °C for 1 h. After regular workup, the resulting crude product was purified by flash chromatography (EtOAc/hexanes = 1/2) to afford compound **11** (200 mg, 80%) as a gel:  $[\alpha]^{24}_D = +28.7$  (*c* 0.90, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 1.38 (m, 3H), 2.27 (s, 3H), 4.19 (t, *J* = 5.7 Hz, 1H), 4.36–4.42 (m, 3H), 5.13 (m, 1H), 5.22 (dd, *J* = 12.3, 7.9 Hz, 1H), 5.28 (d, *J* = 5.7 Hz, 1H), 5.59 (dd, *J* = 17.7, 7.5 Hz, 1H), 7.26–7.30 (m, 8H), 7.36 (t, *J* = 7.0 Hz, 2H), 7.56 (m, 2H), 7.73 (d, *J* = 7.5 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 18.6, 27.9, 47.0, 50.1, 63.1, 67.0, 68.2, 119.9, 125.0, 127.0, 127.6, 128.3, 128.4 (2 C), 128.6 (2 C), 128.7, 128.8, 134.4, 141.2, 143.6, 143.8, 155.9, 165.6, 172.3, 197.9; HRMS (MALDI-FTMS) calcd for C<sub>29</sub>H<sub>28</sub>N<sub>2</sub>O<sub>6</sub> (M + Na<sup>+</sup>) 523.1839, found 523.1836.

**Compound 4.** Compound **4** was synthesized from **11** in 84% yield as a colorless gel following the procedure used for the synthesis of **8**:  $[\alpha]^{24}_D = +26.2$  (*c* 0.81, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 1.54 (d, *J* = 6.6 Hz, 3H), 2.53 (s, 3H), 4.20 (t, *J* = 7.0 Hz, 1H), 4.40 (d, *J* = 6.6 Hz, 2H), 5.00 (m, 1H), 5.35 (s, 2H), 5.59 (d, *J* = 7.4 Hz, 1H), 7.28–7.39 (m, 7H), 7.42 (d, *J* = 7.0 Hz, 2H), 7.58 (t, *J* = 7.9 Hz, 2H), 7.75 (d, *J* = 7.5 Hz, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 12.1, 20.2, 45.1, 47.1, 66.6, 67.0, 119.9, 125.0, 127.0, 127.4, 127.6, 128.3, 128.4, 128.5, 135.6, 141.2, 143.7, 143.8, 155.5, 156.6, 162.5, 168.9; HRMS (MALDI-FTMS) calcd for C<sub>29</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub> (M + Na<sup>+</sup>) 483.1914, found 483.1916; HPLC (chiralcel OD-H column, 254 nm, 90/10 hexane/2-propanol to 70/30 hexane/2-propanol gradient over 60 min, flow = 1.0 mL/min) *t*<sub>R</sub> = 30.9 (*R*), 39.8 (*S*) min.

**Compound 12.** Pd(OAc)<sub>2</sub> (4.5 mg, 0.02 mmol) and styrene polymer-bound triphenylphosphine (101 mg, 1.59 mol/g, 0.16 mmol) were added to a flask containing CH<sub>2</sub>Cl<sub>2</sub> (10 mL). After the mixture was stirred for 10 min, **3** (462 mg, 1 mmol) and PhSiH<sub>3</sub> (0.24 mL, 2 mmol) were added separately. The reaction progress was monitored by TLC, and the reaction was complete in 15 min. After removal of the solvent, the residue was passed through a short silica gel column eluted with CHCl<sub>3</sub>/EtOH (1/1). The carboxylic acid was used in the next step without further purification. Coupling with **2** (391 mg, 0.9 mmol) following the procedure used for the synthesis of **6** gave **12** as a white foam (511 mg, 92%):  $[\alpha]^{24}_D = +23.6$  (*c* 0.74, CHCl<sub>3</sub>); <sup>1</sup>H

NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 0.93 (d, *J* = 6.6 Hz, 3H), 0.95 (d, *J* = 6.6 Hz, 3H), 1.79 (d, *J* = 6.5 Hz, 3H), 2.28 (m, 1H), 4.22 (m, 1H), 4.47 (d, *J* = 6.6 Hz, 2H), 4.81 (d, *J* = 5.7 Hz, 2H), 4.87 (dd, *J* = 8.3, 6.6 Hz, 1H), 5.27 (d, *J* = 10.5 Hz, 1H), 5.38 (d, *J* = 17.1 Hz, 1H), 5.60 (m, 1H), 5.73 (d, *J* = 8.8 Hz, 1H), 6.01 (m, 1H), 7.25–7.29 (m, 2H), 7.38 (dd, *J* = 7.5, 7.0 Hz, 2H), 7.59 (t, *J* = 8.4 Hz, 2H), 7.75 (d, *J* = 7.5 Hz, 2H), 8.02 (s, 1H), 8.04 (d, *J* = 8.3 Hz, 1H), 8.10 (s, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 17.6, 19.4, 20.9, 33.0, 47.1 (2 C), 58.4, 65.8, 66.7, 118.8, 119.9, 123.6, 124.9, 126.9, 127.6 (2 C), 131.7, 141.2, 143.6, 146.6, 149.2, 156.0, 160.4, 160.7, 171.9, 173.0; HRMS (MALDI-FTMS) calcd for C<sub>32</sub>H<sub>32</sub>N<sub>4</sub>O<sub>5</sub>S<sub>2</sub> (M + H<sup>+</sup>) 617.1887, found 617.1902.

**Compound 13.** Pd on activated carbon (15 mg) was added to a flask containing **4** (154 mg, 0.32 mmol) in MeOH (3 mL) and THF (3 mL). The reaction flask was filled with a H<sub>2</sub> balloon, evacuated, and purged with H<sub>2</sub> three times. The reaction progress was monitored by TLC and was complete in 1 h. After removal of the solvent, the residue was passed through a short silica gel column eluted with MeOH. The carboxylic acid was used in the next step without further purification. By following the procedure for the synthesis of **6**, the carboxylic acid was coupled with **12** (185 mg, 0.3 mmol) affording **13** as a white foam (217 mg, 94%):  $[\alpha]^{24}_D = +30.7$  (*c* 1.46, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 1.01 (d, *J* = 6.1 Hz, 3H), 1.02 (d, *J* = 6.1 Hz, 3H), 1.59 (d, *J* = 6.6 Hz, 3H), 1.79 (d, *J* = 7.0 Hz, 3H), 2.45 (m, 1H), 2.61 (s, 3H), 4.22 (dd, *J* = 7.0, 6.6 Hz, 1H), 4.42–4.44 (m, 2H), 4.82 (d, *J* = 5.7 Hz, 2H), 5.02 (m, 1H), 5.25–5.85 (m, 2H), 5.38 (d, *J* = 17.1 Hz, 1H), 5.56–5.61 (m, 2H), 6.00 (m, 1H), 7.27–7.29 (m, 2H), 7.38 (t, *J* = 7.5 Hz, 2H), 7.47 (d, *J* = 9.2 Hz, 1H), 7.58 (dd, *J* = 8.7, 8.3 Hz, 2H), 7.75 (d, *J* = 7.5 Hz, 2H), 7.91 (d, *J* = 7.9 Hz, 1H), 8.01 (s, 1H), 8.08 (s, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 11.7, 17.9, 19.5, 19.6, 20.9, 33.0, 45.0, 47.0 (2 C), 55.9, 65.9, 66.9, 118.8, 119.9, 123.6, 124.9, 126.9, 127.7, 128.4, 131.7, 141.2, 143.5, 143.7, 146.6, 149.1, 154.0, 155.5, 160.4, 160.8, 161.3, 161.5, 171.8, 173.1; HRMS (MALDI-FTMS) calcd for C<sub>39</sub>H<sub>40</sub>N<sub>6</sub>O<sub>7</sub>S<sub>2</sub> (M + H<sup>+</sup>) 769.2473, found 769.2479.

**Dendramide A (1).** Deprotecting the Fmoc group in **13** (76.9 mg, 0.1 mmol) used the procedure described for the synthesis of **6**. The allyl group was removed by a palladium catalyst as described in the synthesis of **12**. The resulting amino acid **14** was dissolved in CH<sub>2</sub>Cl<sub>2</sub>/DMF (10 mL, v/v: 2/1) as a 0.01 M solution. This solution was added to a flask containing PyBOP (104 mg, 0.2 mmol) and DMAP (24.4 mg, 0.2 mmol) in CH<sub>2</sub>Cl<sub>2</sub>/DMF (20 mL, v/v 2/1) over 8 h using a syringe pump. After the addition was complete, the mixture was stirred for 2 h. Regular workup and purification by flash chromatography gave **1** as a colorless semisolid (39.6 mg, 81%):  $[\alpha]^{25}_D = +83.8$  (*c* 0.76, CHCl<sub>3</sub>) [lit.<sup>3b</sup>  $[\alpha]_D = +53.9$  (*c* 0.2, CHCl<sub>3</sub>)]; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ 0.98 (d, *J* = 7.0 Hz, 3H), 1.08 (d, *J* = 7.0 Hz, 3H), 1.71 (d, *J* = 7.0 Hz, 3H), 1.73 (d, *J* = 7.0 Hz, 3H), 2.33 (m, 1H), 2.68 (s, 3H), 5.21 (q, *J* = 6.6 Hz, 1H), 5.31 (dd, *J* = 7.9, 4.8 Hz, 1H), 5.72 (dq, *J* = 8.3, 6.6 Hz, 1H), 8.14 (s, 1H), 8.15 (s, 1H), 8.49 (d, *J* = 8.3 Hz, 1H), 8.55 (d, *J* = 8.3 Hz, 1H), 8.65 (d, *J* = 6.1 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 11.6, 18.3, 18.4, 20.9, 24.9, 35.1, 44.3, 47.0, 55.9, 123.7, 123.8, 128.4, 148.8 (2 C), 153.8, 159.6, 159.8, 160.5, 161.7, 168.2, 171.1; HRMS (MALDI-FTMS) calcd for C<sub>21</sub>H<sub>24</sub>N<sub>6</sub>O<sub>4</sub>S<sub>2</sub> (M + H<sup>+</sup>) 489.1373, found 489.1373.

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**Supporting Information Available:** General procedures and NMR spectra for **1–13**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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